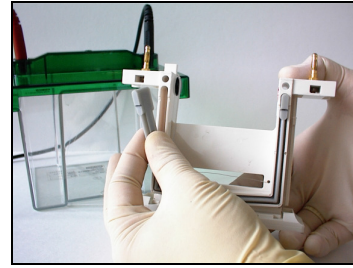


# Special Instructions for Mini Snap-A-Gel™ Universal Cassettes

## Bio-Rad Mini Protean II™ & III™

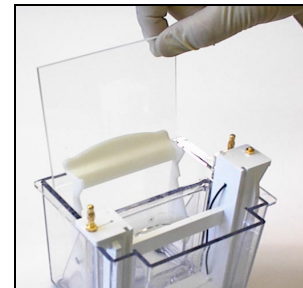
- Remove the inner frame from the apparatus.
- Gently pull the gasket out using your fingers (see right). Clean the groove and the gasket using a mild detergent.
- Dry the groove and gasket. *Reinsert the gasket so the smooth side of the gasket faces outward.*
- The smooth face of the Jule Cassettes against the smooth gasket will prevent upper buffer tank leakage.
- **For BioRad Protean II users, Turn each of the 4 screws ¼ turn after screw contacts cassette. Do not over tighten.**



## Novex Xcell™ I, II, & SureLock™

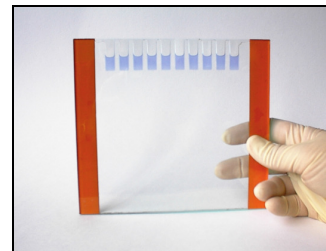
- For adequate sealing pressure, place the supplied *plastic spacer* into the apparatus at the back of the tension wedge (SureLock unit) or between the wedge and rear cassette (Xcell units).

**Note:** The Snap-A-Gel cassette provides the maximum path length in an 8x10cm cassette. Jule's cassette will sit lower in your apparatus and require less buffer. Fill the upper buffer chamber to approximately 0.5 cm above the short plate. Add 300 ml of buffer to the lower buffer chamber. The gel's path length is comparable to many 10cm x 10cm cassettes.



## Large 1-D and 2-D Gels

- Remove gel from its sealed pack.
- Assemble into your apparatus according to the manufacturer's instructions.
- Fill upper buffer chamber and check for leaks.
- Fill lower buffer chamber and load samples.
- Connect apparatus to power supply. **Run gel(s) at constant voltage.**
- Begin run using "Starting Current" from table below. After 90 minutes, reduce current according to table below. Run at the voltage which yields desired current
- When finished, remove gel for staining or blotting.



Recommended Power Settings					Sample Loading		
<i>Run Gels at Constant Voltage</i>							
Gel Type	Thickness	Starting Current Milliamps per Gel	Reduce Current After 90 mins Milliamps per Gel	Run Time (hours)	Gel Type	Total Well Volume	Recommended Sample Load
Tris-Glycine	0.75 mm	30	18	4-6	0.75 mm		
	1.00 mm	40	24	4-6	10 well	110 µL	10 - 105 µL
	1.50 mm	50	30	4-6	15 well	80 µL	10 - 75 µL
TBE	0.75 mm	25	15	4-6	1.00 mm		
	1.00 mm	30	18	4-6	10 well	145 µL	10 - 140 µL
	1.50 mm	35	20	4-6	15 well	110 µL	10 - 105 µL
Tricine	0.75 mm	25 - 35	15 - 20	8-12	1.50 mm		
	1.00 mm	35 - 50	20 - 30	8-12	10 well	220 µL	10 - 215 µL
	1.50 mm	50 - 65	30 - 40	8-12	15 well	165 µL	10 - 160 µL

# Precast Snap-A-Gel™ Quick Guide

**\*\* Bio-Rad and Novex Users \*\*  
Special Instructions on Reverse Side**

For Your Safety: Always wear eye protection and latex gloves.

## Mini Gels (Large Gels see Page 2)

Snap Off the Bottom Tab *Before* Running Gel:

- Cut foil pack and remove gel.
- Place cassette on a lab bench with the bottom tab overhanging the edge and the notched plate facing up.
- Holding the cassette down with one hand rotate the tab down, then up. Use pliers for added leverage.



Electrophoresis:

- Remove comb and rinse the sample wells with 1X running buffer.
- *For easy sample loading* - Wet and apply the enclosed Snappy-Load™ Decal to the back of the cassette.
- Place cassette in your apparatus according to its Operator's Manual
- Fill the upper buffer chamber and check for leaks.
- Load your sample into the wells and fill the lower buffer chamber
- Load all unused wells with sample buffer
- Connect the apparatus to the power supply. **Run gel(s) at constant voltage.**

*Refer to Jule, Inc.'s recommended sample loading and power settings and the Operator's Manual for your electrophoresis apparatus.*

Recommended Power Settings <i>Run Gels at Constant Voltage</i>				Sample Loading		
Gel Type	Thickness	Starting Current		Gel Type	Total Well Volume	Recommended Sample Load
		Milliamps per Gel	Run Time			
Tris-Glycine	0.75 mm	30	45 – 60 min.	0.75 mm		
	1.50 mm	35	45 – 60 min.	10 well	25 µL	2-15 µL
TBE	0.75 mm	15	60 min.	15 well	18 µL	2-10 µL
	1.50 mm	20	60 min.	1.50 mm		
Tricine	0.75 mm	30 – 40	60 – 90 min.	10 well	50 µL	3-30 µL
	1.50mm	55 – 60	60 – 90 min.	15 well	37 µL	3-20 µL

Remove Gel from the Cassette:

- Place cassette on the lab bench as shown.
- Insert the tip of a small screwdriver between the plates near the bottom of the cassette.
- Twist the screwdriver gently and listen for a “snap”. Moving along the cassette's edge, repeat until the entire seal is broken.
- Break the seal on the opposite side the same way
- Carefully lift the top plate and remove the gel for staining or blotting.

